



**Improvement of Growth Rate in *In Vitro* Culture of *Paphiopedilum primulinum* M. W. Wood & P. Taylor and *Paphiopedilum glaucophyllum* J. J. Smith using Banana Enrichment Media**

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**Highlights**

- The impact of supplementing the medium with banana homogenate on the orchid species *Paphiopedilum primulinum* and *Paphiopedilum glaucophyllum* was determined by comparing the plants' heights (measured from the base to the tip of the stem), leaf count and root count.
- ½ Murashige and Skoog (MS) + *Ambon Lumut* banana (ALB) homogenate is a better medium for *P. primulinum* and *P. glaucophyllum* growth than media without banana homogenate.
- Culture medium added banana homogenate was able to support the propagation of plants.

## **Improvement of Growth Rate in *In Vitro* Culture of *Paphiopedilum primulinum* M. W. Wood & P. Taylor and *Paphiopedilum glaucophyllum* J. J. Smith using Banana Enrichment Media**

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**Abstract:** *Paphiopedilum primulinum* and *Paphiopedilum glaucophyllum* have unique labellum colour and shaped like lady's slippers. These orchids are from the Cochlopetalum section, which is exclusively found in Sumatra and Java. There are so many people that desire to collect these plants illegally. Due to extensive commercial exploitation, *Paphiopedilum* is in danger of going extinct. Tissue culture techniques are utilised to conserve threatened orchid germplasm in a short time. The success of the *in vitro* culture depends on the accuracy of the basic media composition used. The Ambon Lumut banana (ALB) can accelerate plant growth and cell division. Banana added to the culture medium was prepared by mashing the ripe flesh (3.5 months old) using a mortar. This research aims to investigate the effect of banana homogenate supplemented media for the orchids *P. primulinum* and *P. glaucophyllum* based on the parameters of difference of plant height (calculated from the base of the stem to the tip of the plant stem), number of leaves, and number of roots. The measurement method was carried out using a ruler with a centimetre scale. Observations and documentation were carried out once a week for 7 weeks after planting (WAP) for *P. primulinum* and *P. glaucophyllum*. The results showed that ½ Murashige and Skoog (MS) + ALB homogenate is a better medium for *P. primulinum* and *P. glaucophyllum* growth than media without banana homogenate. The highest values of plant height, leaf growth and root growth of *P. primulinum* with banana homogenate were 0.44 cm, 0.63 leaves, and 0.50 roots, respectively. The highest values of plant height and leaf growth of *P. glaucophyllum* were 0.75 cm and 1.90 leaves, respectively. Culture medium added banana homogenate was able to support the propagation of plants, some of which

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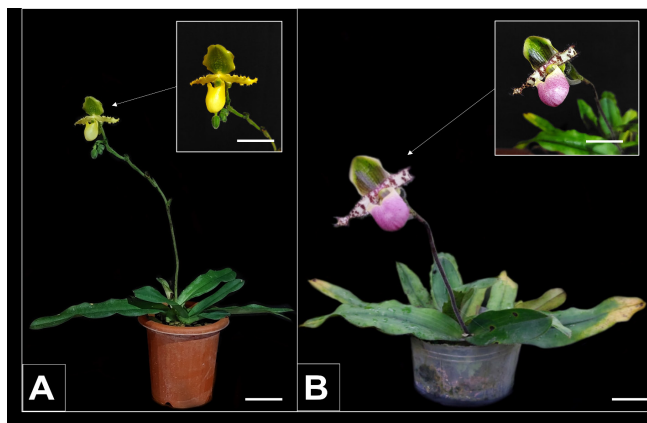
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are returned to nature and others used for industrial purposes (conventionally cultivated by the community).

**Keywords:** *In vitro* Culture, *Paphiopedilum primulinum*, *Paphiopedilum glaucophyllum*, Treatment Media

## INTRODUCTION

Orchids are ornamental plants that have high economic value because their morphology and colour are diverse and attractive (Luo *et al.* 2021). One of the orchid genera that has a unique pouch-shaped lip of a lady's slipper-like labellum is the genus *Paphiopedilum*, which is included in the Appendix 1 category based on the Convention of International Trades in Endangered Species of Wild Fauna and Flora (CITES). The genus *Paphiopedilum* is the most widely cultivated and hybridised genus of orchids. Overexploitation or illegal activities, natural disasters, conversion of land into settlements and habitat destruction are the causes of *Paphiopedilum* population decline in nature (Kaur & Bhutani 2013; Sindiya *et al.* 2018). According to Azmi and Wiendi (2015), it takes a long time to produce *Paphiopedilum* seeds in large quantities. This plant reproduces by producing saplings, and it takes two to three years to produce saplings (Birk 1983). Two of the endangered *Paphiopedilum* species are *Paphiopedilum primulinum* and *P. glaucophyllum*. *P. primulinum* is a pouch-shaped lip of a lady's slipper orchid endemic to Sumatra (southern Aceh), and the abundance of this orchid species has decreased significantly in the last few decades (Fig. 1A). In addition, another *Paphiopedilum* species that is threatened with extinction is *P. glaucophyllum* J. J. Smith, which is a plant native to West Java that was discovered and introduced by J. J. Smith in 1900 (Fig. 1B) (Azmi & Wiendi 2015; Destri & Ismaini 2018).



**Figure 1:** Habitus of (A) *P. primulinum* and (B) *P. glaucophyllum*.

Note: Bars = 5 cm.

Tissue culture techniques are used to conserve rare and endangered orchid germplasm (Pant *et al.* 2022). The success of *in vitro* culture is influenced by the growth media and growth effect substances (Plant growth regulators [PGRs]) used (Chaudhary & Prakash 2019). Tissue culture media generally contain macro- and micronutrients, vitamins, amino acids, sucrose and growth regulators. A medium pH that is too low (<4.5) or too high (>7.0) can inhibit or stop the growth and development of *in vitro* cultures (Widiastoety *et al.* 2005). There are several common types of basic *in vitro* culture media, namely, MS – Murashige and Skoog (used on almost all types of plants), VW – Vacin and Went, NP – New Phalaenopsis and KC – Knudson C. Organic substances can enhance the process of development and regeneration of *in vitro* plants (Utami & Hariyanto 2020).

*Ambon Lumut* banana (ALB) was used because it has higher Ca (7.22 mg), and vitamin C (19.10 mg) content than *Ambon* and *Raja* bananas. Bananas are often added to *in vitro* culture media, and the addition of 25 g/L ALB into the culture medium stimulated the multiplication of *Phalaenopsis fuscata* shoots and leaves (Garvita & Handini 2011). Nurfadilah *et al.* (2018) reported that ALB extract had the highest glucose content compared to other types of bananas. Glucose can be used as an energy source to stimulate cell division and promote cell differentiation, as well as trigger shoot growth. There are no studies yet of plantlet growth of *P. primulinum* and *P. glaucophyllum* orchids in relation to the addition of banana extract into culture media. Therefore, it is necessary to research the growth of *P. primulinum* and *P. glaucophyllum* orchids cultured in various types and concentrations of media ( $\frac{1}{2}$  MS,  $\frac{1}{2}$  MS + ALB, MS,  $\frac{1}{2}$  NP, NP, KC and VW) *in vitro*.

## MATERIALS AND METHODS

### Plant Materials, Growth Conditions and Culture Conditions

The plant materials used in this research were 1-year-old *P. primulinum* and 2-year-old *P. glaucophyllum* orchid plantlets from the Research Center for Plant Conservation and Botanical Gardens, Indonesian Institute of Sciences BRIN (in-kind support used in this research). Plantlets were cultured on  $\frac{1}{2}$  MS,  $\frac{1}{2}$  MS + ALB, MS,  $\frac{1}{2}$  NP, NP, KC and VW treatment media (Merck Darmstadt, Germany) without the addition of growth regulators (PGR). The bananas used are ripe banana flesh that are about 3.5 months old. Banana homogenate was prepared by weighing the peeled banana at 20 g/L. The banana was mashed using a blender or mortar, added to  $\frac{1}{2}$  MS media and homogenised. Plantlets were planted in culture bottles containing treatment media using sterile tweezers and then placed in petri dishes to be cleaned of residual media. Cultures were maintained in treatment of light for 24 h and at a temperature of 25°C with 70% humidity.

### **Nutritional assessment of ALB**

The nutritional content of ALB was assessed using the Atomic Absorption Spectroscopy (AAS) test method at the Laboratory of Food and Agricultural Product Technology, Faculty of Agricultural Technology, Universitas Gadjah Mada and the Integrated Research and Testing Laboratory, Universitas Gadjah Mada.

### **Measurement of Height and Leaf and Root Growth of *P. primulinum* and *P. glaucophyllum***

The parameters observed in this study were plant height (measured from the base of the stem to the tip of the main stem), the number of leaves, and the number of roots growing on each plantlet. The tool used to measure plantlet height is a ruler with a centimetre scale. Growth was observed and documented once a week. Observations were made for 7 weeks after planting (WAP) for *P. primulinum* and *P. glaucophyllum*.

### **Research Design**

Due to the limited number of plants available as samples—considering that the plants included were uncommon species classified as endangered—only three repetitions of the experiment were conducted. In the experiments, three bottles were used, and each bottle contained one plantlet (each plantlet was referred to as one replicate). The design used in this study was a completely randomised design that was analysed by statistical ANOVA test through SPSS IBM Statistics 25. If there were differences, the test was continued with Duncan's multiple distance test 5%.

## **RESULTS AND DISCUSSION**

### **The Growth Response of *P. primulinum* and *P. glaucophyllum* Plantlets**

The results showed that *P. primulinum* and *P. glaucophyllum* plantlets cultured on various types and concentrations of treatment media showed different growth parameters for plant height, leaf growth, and root growth. Based on ANOVA, which was further analysed using Duncan's 5% test, the increases in plant height, leaf growth, and root growth were significantly different. This is shown in Table 1.

**Table 1:** Average results of observations of plant height, leaf growth, and root growth on *P. primulinum* and *P. glaucophyllum* on various types and concentrations of treatment media.

Media	<i>P. primulinum</i>			<i>P. glaucophyllum</i>		
	Difference of plantlet height (cm)	Number of leaves	Number of roots	Difference of plantlet height (cm)	Number of leaves	Number of roots
½ MS	0.41 ± 0.06 <sup>ab</sup>	0.67 ± 0.13 <sup>a*</sup>	0.42 ± 0.10 <sup>ab</sup>	0.45 ± 0.08 <sup>bc</sup>	0.64 ± 0.10 <sup>bc</sup>	0.42 ± 0.19 <sup>a*</sup>
½ MS+ALB*	0.44 ± 0.07 <sup>a*</sup>	0.63 ± 0.13 <sup>a*</sup>	0.50 ± 0.17 <sup>a*</sup>	0.75 ± 0.08 <sup>a*</sup>	1.9 ± 0.20 <sup>a*</sup>	0.25 ± 0.09 <sup>ab</sup>
MS	0.40 ± 0.09 <sup>abc</sup>	0.67 ± 0.14 <sup>a</sup>	0.17 ± 0.08 <sup>bc</sup>	0.59 ± 0.10 <sup>ab</sup>	0.58 ± 0.20 <sup>bc</sup>	0.21 ± 0.08 <sup>ab</sup>
½ NP	0.26 ± 0.04 <sup>bc</sup>	0.33 ± 0.10 <sup>a</sup>	0.13 ± 0.07 <sup>c</sup>	0.27 ± 0.08 <sup>cd</sup>	0.60 ± 0.10 <sup>bc</sup>	0.12 ± 0.07 <sup>ab</sup>
NP	0.24 ± 0.04 <sup>c</sup>	0.50 ± 0.10 <sup>a</sup>	0.00 ± 0.00 <sup>c</sup>	0.32 ± 0.07 <sup>cd</sup>	1.21 ± 0.40 <sup>b</sup>	0.46 ± 0.17 <sup>a*</sup>
KC	0.26 ± 0.03 <sup>bc</sup>	0.46 ± 0.10 <sup>a</sup>	0.38 ± 0.10 <sup>ab</sup>	0.34 ± 0.06 <sup>cd</sup>	1.26 ± 0.30 <sup>ab</sup>	0.22 ± 0.08 <sup>ab</sup>
VW	0.30 ± 0.04 <sup>abc</sup>	0.33 ± 0.10 <sup>a</sup>	0.13 ± 0.07 <sup>bc</sup>	0.25 ± 0.04 <sup>cd</sup>	0.25 ± 0.10 <sup>c</sup>	0 ± 0.00 <sup>b</sup>

Notes: ½ MS = half-strength MS media; ½ MS + ALB = half-strength MS media with banana homogenate; MS = full-strength MS media; ½ NP = half-strength NP media; NP = full-strength NP media; KC = full-strength KC media; VW = full-strength VW media. The symbol \* represent the highest values from each parameter. Data in the same column followed by the same letters are not significantly different by Duncan's test at  $p \leq 0.05$ .

### Nutrient Value of ALB

Bananas are often used as an organic additive in *in vitro* culture media. They are rich in various natural growth regulators, vitamins, and minerals. One type of banana that is supplemented into media is ALB. The nutritional values of ALB are shown in Table 2.

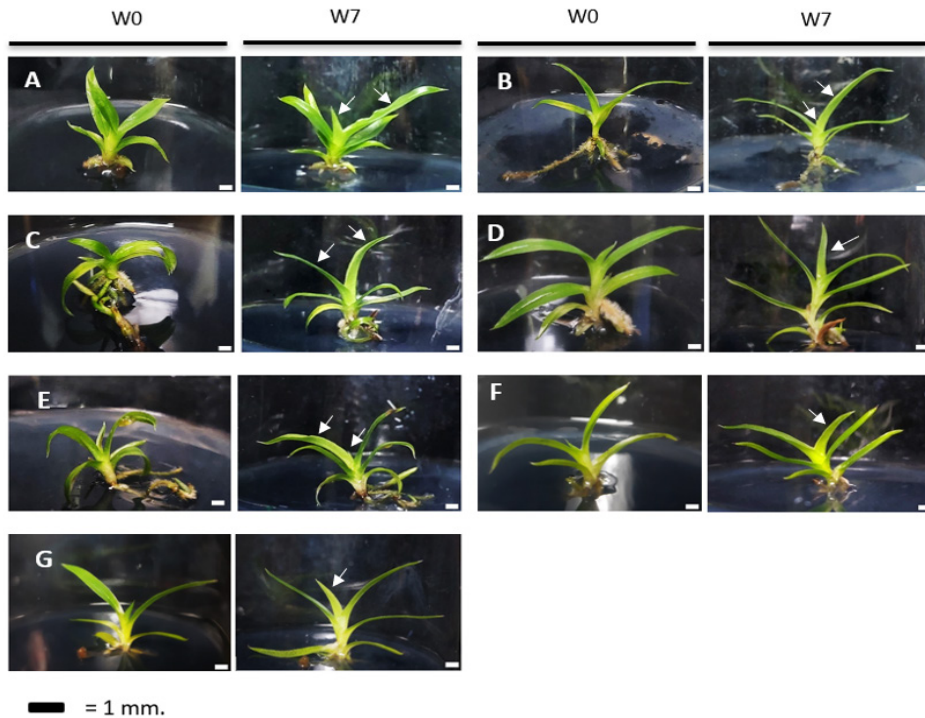
**Table 2:** Nutritional values of ALB in 100 g edible portion.

Nutritional content	ALB <sup>a</sup>	Ambon banana <sup>b</sup>	Raja banana <sup>c</sup>
Water	74.35%	75.70%	66.49%
Protein	0.92%	1.10%	1.51%
Fat	0.19%	0.20%	0.05%
Carbohydrate	23.32%	22.20%	31.13%
Ash	1.28%	0.80%	0.82%
Ca	7.22 mg	7.00 mg	–
Mg	16.25 mg	36.0 mg	–
K	298.9 mg	460.0 mg	350.0 mg
Vitamin C	19.10 mg	10.00 mg	16.45 mg

Notes: a = The content analysis of ALB was conducted by the Test Laboratory of Food and Agricultural Product Technology, Faculty of Agricultural Technology, Universitas Gadjah Mada and the Integrated Research and Testing Laboratory, Universitas Gadjah Mada; b = The nutritional content analysis of Ambon banana is based on Garvita and Handini (2011); c = The nutritional content analysis of Raja banana is based on Hapsari and Lestari (2016).

### Plantlet Development of *P. primulinum* and *P. glaucophyllum*

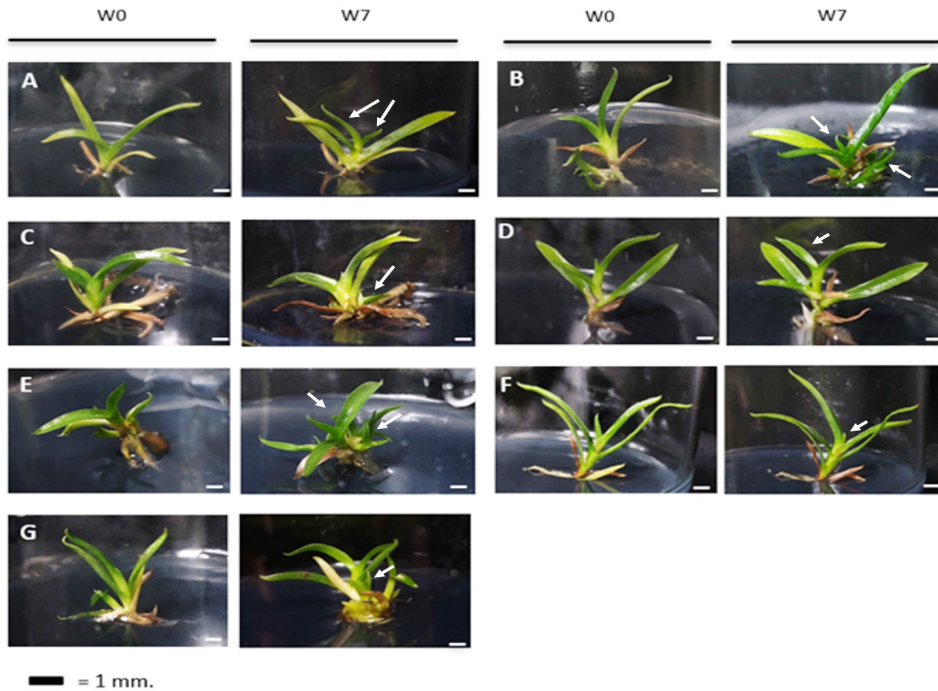
The growth of new leaves is one response that is easily noticeable during the plantlets' development. The growth of leaves on each plantlet, indicated by a white arrow, as seen in Figs. 2 and 3. In every culture medium, both species responded in the same way with the development of new leaves.



**Figure 2:** Plantlet development of *P. primulinum* on various culture media at week 0 and week 7 for (A)  $\frac{1}{2}$  MS, (B)  $\frac{1}{2}$  MS + ALB, (C) MS, (D)  $\frac{1}{2}$  NP, (E) NP, (F) KC, and (G) VW, respectively.

Notes: Bars = 1 cm. The white arrows represent the growth of new leaves.





**Figure 3:** Plantlet development of *P. glaucophyllum* on various culture media at week 0 and week 7 for (A)  $\frac{1}{2}$  MS, (B)  $\frac{1}{2}$  MS + ALB, (C) MS, (D)  $\frac{1}{2}$  NP, (E) NP, (F) KC, and (G) VW, respectively.

Notes: Bars = 1 cm. The white arrows represent the growth of new leaves.

### The Growth Response of Plantlets

The type of media has a significant impact on the growth and development of plantlets (Tuhuteru *et al.* 2012). MS, NP, KC and VW are some types of basic media that are often used in orchid propagation and are available in full-strength and half-strength concentrations. The different contents of macro- and micronutrients in each type of medium have different effects on plant growth (Inkiriwang *et al.* 2016). Based on this study, half-strength MS ( $\frac{1}{2}$  MS) media is the best treatment media; it is a complex media containing macronutrients, micronutrients and vitamins. When the concentration of the media changes, so do the nutrient components. As a result, different media concentrations or strengths can affect plant growth and organogenesis *in vitro* (Rezali *et al.* 2017). MS medium has been used for *in vitro* propagation of several orchid plants, including *Paphiopedilum insigne*, *Vanda pumila* Hook. f., *Tolumnia Louise Elmore 'Elsa'* (Deb & Jakha 2020). NP media was used to culture *Dendrobium lineale*, *Dendrobium phalaenopsis* and *Phalaenopsis amabilis* (Mustika & Semiarti 2021; Mose *et al.* 2020; Zulwanis *et al.* 2020).

The plant height of *P. glaucophyllum* was higher than that of *P. primulinum*, which is due to the different endogenous hormones contained in each species.



The results of this study showed the highest leaf growth of *P. primulinum* in ½ MS media (0.67) and the highest leaf growth of *P. glaucophyllum* in ½ MS + ALB (1.90). The number of leaves formed on each planted explant is controlled by the balance and interaction between growth regulators both contained in the explant itself (endogenous) and absorbed from the media (exogenous).

Bhojwani and Razdan (1996) reported that ½ MS or ¼ MS medium produced better results in *Dendrocalamus* shoot propagation than full-strength MS medium. Furthermore, ½ MS medium is effective for propagating sugarcane orchids (*Grammatophyllum speciosum*), initiating shoot and root formation in *Phalaenopsis* protocorm-like bodies (PLB), and supporting shoot and root proliferation in the rare orchid *V. pumila* (Kriswanto et al. 2017; Maharjan et al. 2019; Sulichantini et al. 2021). In the previous study by Kaur and Bhutani (2012), *in vitro* propagation of *Cymbidium pendulum* supplemented with 50 g/L banana homogenate showed the highest number of induced shoots. Banana extract has potential as a sucrose replacement, and the sugar concentration in banana extract is high (27%). The addition of banana extract into the culture medium was able to produce the highest values for shoot induction and shoot height (Phillips et al. 2021). The banana homogenate contains a high concentration of potassium (K), which is essential to support the shoot regeneration of cultured plantlets (Lui et al. 2022).

Aside from having a positive effect on plant height, half-strength MS supplemented banana homogenate (½ MS + ALB) had the highest value of root growth on *P. primulinum*, with a value of 0.50 (Table 1). The highest value of root growth in *P. glaucophyllum* plantlets was shown in ½ MS and NP media. NP is a medium designed specifically for orchid germination that contains high nitrogen content that promote root initiation (Islam et al. 1998; Krisdianto et al. 2020). Because of the high nitrogen content in NP media, the media has a very positive effect on the root growth of *P. glaucophyllum*. Zeng et al. (2013) reported that ½ MS medium was more efficient for rooting *Withania coagulans* because its osmotic strength was lower than full-strength MS. Kartikaningrum et al. (2017) reported that MS media with complete nutrients causes browning in some orchid species such as *Rhyncostilis retusa*, *Dendrobium anosmum* and *Renanthera matutina* when cultured in the media.

Bananas contain a high concentration of carbohydrates and K, which are used as a source of sugar and macronutrients. *Ambon* banana and ALB are two types of bananas that are frequently used as supplements to *in vitro* growth media (Garvita & Handini 2011; Sulichantini et al. 2021). The glucose content of *Ambon* banana can stimulate shoot growth and cell division processes (Nurfadilah et al. 2018). Elements such as K, iron (Fe), and phosphorus (P) are also important for growth, so the ½ MS media treatment with banana pulp shows significant results. Table 2 shows the ALB content. ALB provided 298.8 mg of K per 100 g. Ranjha et al. (2022) reported that banana is an important source of K. A normal-sized banana can provide 350 mg of K. Each 100 g of *Musa acuminata* Colla provides 358 mg of K. In addition, 100 g of *Musa acuminata* Colla provides 5 mg of calcium (Ca) and 27 mg of magnesium (Mg). Ca is related to certain processes, such as

membrane structure and function, ion uptake, reactions with growth regulators and enzymatic activation. The structural function of Ca is characterised by its use in the synthesis of new cell walls, especially the middle lamella, which separates newly dividing cells (Malavolta *et al.* 1997; Taiz & Zeiger 2006). Ca plays an important role in the transmission of hormonal signals; in media supplemented with Ca, the process of mitosis-cytokinesis increases. Ca is also able to stabilise cell membranes, plays a role in cell elongation and cell division, and affects cell pH.

Kaewubon and Meesawat (2014) showed that MS media supplemented with 50 g/L banana pulp can stimulate the development of *Paphiopedilum niveum* PLB into organised plant structures. *Ambon* banana contains hormones in the form of auxin and cytokinin, which can stimulate cell division and differentiation, which is beneficial for cell multiplication (Nurfadilah *et al.* 2018). Half-strength MS media supplemented banana pulp can increase the vitamin content of the media, which supports better growth. Thiamine, pyridoxine and nicotinic acid are three vitamins that are commonly used in plant *in vitro* culture (Hapsoro *et al.* 2018; Hasanah *et al.* 2014). Thiamine (vitamin B1) acts as a coenzyme in many metabolic pathways, particularly those involved in energy production and central metabolism, such as carbon assimilation and respiration (Fitzpatrick & Chapman 2020). According to Garvita and Handini (2011), banana extract contains thiamine, which plays a role in accelerating cell division of the root meristem. *Ambon* bananas, which are equivalent to Cavendish bananas, have thiamine levels of 0.04 mg/100 g (Garvita & Handini 2011; Sallolo *et al.* 2012). Thiamine at high concentrations promotes plant proliferation by inhibiting oxidative stress reactions that can damage plant tissue. This is the reason ½ MS + ALB media resulted in the greatest increase in plant height and root growth when compared to ½ MS media.

## CONCLUSION

Half-strength medium supplemented ALB (½ MS + ALB) was the best medium for the growth of *P. primulinum* orchids based on the parameters of plant height and root growth and the best medium for *P. glaucophyllum* orchids based on the parameters of plant height and leaf growth.

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## AUTHORS' CONTRIBUTIONS

Safitri: Data analysis, writing and editing.  
Dyah Carinae Yalapuspita: Data analysis, writing and editing.  
Elizabeth Handini: Writing and editing.  
Popi Aprilianti: Writing and editing.  
Yupi Isnaini: Writing and editing.  
Endang Semiarti: Conceptualisation, validation, project administration, funding, writing, editing and supervision.

## REFERENCES

- Azmi T K K and Wiendi N M A. (2015). Perbanyakannya anggrek spesies *Paphiopedilum glaucophyllum* J. J. Smith melalui proliferasi tunas adventif secara *in vitro*. *Jurnal Hortikultura Indonesia* 4(3): 115–123. <https://doi.org/10.29244/jhi.4.3.115-123>
- Bhojwani S S and Razdan M K. (1996). Plant tissue culture: Theory and practice. In S S Bhojwani and M K Razdan (eds.), *Studies in plant science*, vol. 5. Amsterdam: Elsevier. [https://doi.org/10.1016/S0928-3420\(96\)80002-4](https://doi.org/10.1016/S0928-3420(96)80002-4)
- Birk L A. (1983). *The Paphiopedilum grower's manual*. Santa Barbara, CA: Pisang Press.
- Chaudhary K and Prakash J. (2019). Effect of 2, 4-D and picloram on somatic embryogenesis in *Carica papaya* var. P-7-9. *Plant Tissue Culture and Biotechnology* 29(1): 25–32. <https://doi.org/10.3329/ptcb.v29i1.41976>
- Deb C R and Jakha H Y. (2020). Factors affecting asymbiotic immature seed culture and *in vitro* propagation of *Paphiopedilum insigne* (Wall. Ex. Lindl.) Pfitzer – A horticultural important vulnerable orchid. *Plant Cell Biotechnology and Molecular Biology* 21(15&16): 129–141.
- Destri D and Ismaini L. (2018). *Paphiopedilum* spp. (Orchidaceae) collection at Cibodas Botanic Garden. In: *3rd International Conference on Tropical Biology "Conservation, Enhancement and Sustainable Use of Indigenous Tropical Flora and Fauna,"* EAMEO BIOTROP, Bogor-Indonesia, 20–21 September, 88–93.
- Fitzpatrick T B and Chapman L M. (2020). The importance of thiamine (vitamin B1) in plant health: From crop yield to biofortification. *Journal of Biological Chemistry* 295(34): 12002–12013. <https://doi.org/10.1074/jbc.REV120.010918>
- Garvita R V and Handini E. (2011). Pengaruh penambahan berbagai kadar pisang dan ubi jalar pada pertumbuhan kultur tiga jenis phalaenopsis [The influence of banana and sweet potato on the development of three phalaenopsis cultures]. *Bulletin Kebun Raya* 14(2): 9–18.
- Hapsari L and Lestari D A. (2016). Fruit characteristic and nutrient values of four Indonesian banana cultivars (*Musa* spp.) at different genomic groups. *Agrivita Journal of Agricultural Science* 38: 303–311. <https://doi.org/10.17503/agrivita.v38i3.696>
- Hapsoro D, Septiana V A, Ramadiana S and Yusnita. (2018). A medium containing commercial foliar fertilizer and some organic additives could substitute MS medium for *in vitro* growth of *Dendrobium* hybrid seedlings. *Jurnal Floratek* 13(1): 11–22.
- Hasanah U, Rahayu E S and Sumadi. (2014). Pemanfaatan pupuk daun, air kelapa dan bubur pisang sebagai komponen medium pertumbuhan plantlet anggrek *Dendrobium kelemense*. *Biosaintifika: Journal of Biology and Biology Education* 6(2): 161–168.

- Inkiriwang A E B, Mandang J and Runtunuwu S. (2016). Substitusi media Murashige dan Skoog/MS dengan air kelapa dan pupuk daun majemuk pada pertumbuhan anggrek *Dendrobium* secara in vitro [In vitro growth of *Dendrobium* orchids under substitution Murashige dan Skoog/MS medium with coconut water and compound leaf fertilizer]. *Jurnal Bios Logos* 6(1): 15-19. <https://doi.org/10.35799/jbl.6.1.2016.16258>
- Islam M O, Ichihashi S and Matsui S. (1998). Control of growth and development of protocorm like body derived from callus by carbon sources in *Phalaenopsis*. *Plant Biotechnology* 15(4): 183–187. <https://doi.org/10.5511/plantbiotechnology.15.183>
- Kaewubon P and Meesawat U. (2014). Histological analysis of somatic embryogenesis induced in tropical lady's slipper orchid (*Paphiopedilum niveum*). *Acta Horticulturae* 1025: 231–236. <https://doi.org/10.17660/ActaHortic.2014.1025.33>
- Kartikaningrum S, Pramanik D, Dewanti M, Soehendi R and Yufdy M P. (2017). Konservasi anggrek spesies alam menggunakan eksplan biji pada media Vacin & Went [Conservation of orchid natural species using seed explants on Vacin & Went medium]. *Buletin Plasma Nutfah* 23(2): 109–118. <https://doi.org/10.21082/blpn.v23n2.2017.p109-118>
- Kaur S and Bhutani K K. (2012). Organic growth supplement stimulants for in vitro multiplication of *Cymbidium pendulum* (Roxb.) Sw. *Horticultural Science* 39(1): 47–52. <https://doi.org/10.17221/52/2011-hortsci>
- \_\_\_\_\_. (2013). In vitro propagation of *Paphiopedilum spicerianum* (Reichb. F.) Pfitz. *Floriculture and Ornamental Biotechnology* 7(1): 65–70.
- Krisdianto A, Saptiningsih E, Nurchayati Y and Setiari N. (2020). Growth of *Phalaenopsis amabilis* (L.) Blume orchid plantlet on subculture stage by difference of media types and pepton concentrations. *Metamorfosa: Journal of Biological Sciences* 7(2): 182–190. <https://doi.org/10.24843/metamorfosa.2020.v07.i02.p06>
- Kriswanto B, Soeparjono S and Restanto D P. (2017). *The propagation of orchid through formation of protocorm like bodies (PLB) for supporting the rescue of Phalaenopsis orchid*. Proceedings of The International Conference of FoSSA, Jember, Indonesia. 1–3 August, 70–74.
- Lui X J, Sriskanda D, Ling W T, Subramaniam S and Chew B L. (2022). The incorporation of coconut water and banana homogenate in the regeneration of fig (*Ficus carica* L.) cv. Violette de Solliès. *Malaysian Applied Biology* 51(5): 13–22. <https://doi.org/10.55230/mabjournal.v51i5.2327>
- Luo B X, Zhang L, Zheng F, Wu K L, Li L, Zhang X H, Ma G H, Teixeira da Silva J A, Fang L and Zeng S J. (2021). Ovule development and in planta transformation of *Paphiopedilum maudiae* by agrobacterium-mediated ovary-injection. *International Journal of Molecular Sciences* 22(1): 84. <https://doi.org/10.3390/ijms22010084>
- Maharjan S, Pradhan S, Thapa B B and Pant B. (2019). In vitro propagation of endangered orchid *Vanda pumila* Hook.f. through protocorms culture. *American Journal of Plant Sciences* 10(7): 1220–1232. <https://doi.org/10.4236/ajps.2019.107087>
- Malavolta E, Vitti G C and Oliveira S A. (1997). *Evaluation of the nutritional status of plants: principles and applications*. Piracicaba, Brazil: POTAFOS.
- Mose W, Daryono B S, Indrianto A, Purwantoro A and Semiarti E. (2020). Direct somatic embryogenesis and regeneration of an Indonesian orchid *Phalaenopsis amabilis* (L.) Blume under a variety of plant growth regulators, light regime, and organic substances. *Jordan Journal of Biological Sciences* 13(4): 509–518.
- Mustika N D and Semiarti E. (2021). In vitro culture of *Dendrobium lineale* Rolfe orchid for plant breeding and propagation. *IOP Conference Series: Earth and Environmental Science* 913: 012066. <https://doi.org/10.1088/1755-1315/913/1/012066>

- Nurfadilah, Mukarlina and Elvi R P W. (2018). Multiplikasi anggrek hitam (*Coelogyne pandurata* Lindl) pada media Murashige Skoog (MS) dengan penambahan ekstrak pisang Ambon dan benzyl amino purin (BAP). *Jurnal Protobiont* 7(3): 47–53. <https://doi.org/10.26418/protobiont.v7i3.29078>
- Pant B, Chand K, Paudel M R, Joshi P R, Thapa B B, Park S Y, Shakya S, Thakuri L S, Rajbahak S, Sah A K, Baniya M K, Gurung P R, Lasta Maharjan L and Rajbhandari P. (2022). Micropropagation, antioxidant and anticancer activity of pineapple orchid: *Dendrobium densiflorum* Lindl. *Journal of Plant Biochemistry and Biotechnology* 31(2): 399–409. <https://doi.org/10.1007/s13562-021-00692-y>
- Phillips K M, McGinty R C, Couture G, Pehrsson P R, McKillop K and Fukagawa N K. (2021). Dietary fiber, starch, and sugars in bananas at different stages of ripeness in the retail market. *PLoS ONE* 16(7): e0253366. <https://doi.org/10.1371/journal.pone.0253366>
- Ranjha M M A N, Irfan S, Nadeem M and Mahmood S. (2022). A comprehensive review on nutritional value, medicinal uses, and processing of banana. *Food Reviews International* 38(2): 199–225. <https://doi.org/10.1080/87559129.2020.1725890>
- Rezali N I, Jaafar Sidik N, Saleh A, Osman N I and Mohd Adam N A. (2017). The effects of different strength of MS media in solid and liquid media on in vitro growth of *Typhonium flagelliforme*. *Asian Pacific Journal of Tropical Biomedicine* 7(2): 151–156. <https://doi.org/10.1016/j.apjtb.2016.11.019>
- Sallolo S T, Sadimantara I G R and Wijayanto T. (2012). Pertumbuhan anggrek *Dendrobium candy* Stripe Lasianthera pada media saph Vacin dan Went secara in vitro dengan penambahan ekstrak pisang raja dan fish emulsion. *Penelitian Agronomi* 1(1): 58–62.
- Sindiya V, Mukarramah L, Rohimah S and Perwitasari D A G. (2018). Studi in silico potensi DNA barcode pada anggrek langka *Paphiopedilum*. *Biosfer* 3(1): 20–26. <https://doi.org/10.23969/biosfer.v3i1.1250>
- Sulichantini E D, Eliyani, Nazari A P D, SusyLOWATI and Saputra A. (2021). Respon pertumbuhan anggrek tebu (*Grammatophyllum speciosum* Blume) secara in vitro terhadap pemberian benzyl amino purin, kinetin, naftalena acetic acid dan ekstrak pisang Ambon dalam media dasar setengah Murashige and Skoog. *Ziraa'Ah Majalah Ilmiah Pertanian* 46(1): 59–69. <https://doi.org/10.31602/zmp.v46i1.3941>
- Taiz L and Zeiger E. (2006). *Plant physiology*, 4th edition. Massachusetts: Sinauer Associates, Inc.
- Tuhuteru S, Hehanussa M L and Raharjo S H T. (2012). Pertumbuhan dan perkembangan anggrek *Dendrobium anosmum* pada media kultur in vitro dengan beberapa konsentrasi air kelapa. *Agrologia: Jurnal Ilmu Budidaya Tanaman* 1(1): 1–12. <https://doi.org/10.30598/a.v1i1.293>
- Utami E S W and Hariyanto S. (2020). Organic compounds: Contents and their role in improving seed germination and protocorm development in orchids. *International Journal of Agronomy* 2020(1): 2795108. <https://doi.org/10.1155/2020/2795108>
- Widiastoety D, Kartikaningrum S and Purbadi. (2005). Pengaruh pH Media terhadap pertumbuhan plantlet anggrek *Dendrobium*. *Jurnal Hortikultura* 15(1): 18–21.
- Zeng S, Wang J, Wu K, Teixeira da Silva J A, Zhang J and Duan J. (2013). In vitro propagation of *Paphiopedilum hangianum* Perner & Gruss. *Scientia Horticulturae* 151: 147–156. <https://doi.org/10.1016/j.scienta.2012.10.032>
- Zulwanis, Setiari N, Gutierrez-Marcos J and Semiarti E. (2020). The expression of AtRKD4 transgene during induction of somatic embryogenesis in transgenic *Dendrobium phalaenopsis* orchid carrying 35S::GR::AtRKD4. *AIP Conference Proceedings* 2260(1): 060015. <https://doi.org/10.1063/5.0015873>